

Blood Collection

Draw Order Guide Included



Green - Lithium Heparin

Must be spun within 2 hours of collection, then refrigerated.

Used for:

Most chemistries EXCEPT for Lithium and IBCT

Required for Troponin, Ammonia, and Lactic Acid



Gold – SST

Must be spun within 2 hours of collection, then refrigerated.

Used for:

Most chemistries EXCEPT Troponin, Ammonia and Lactic Acid



Light Blue – Sodium Citrate

Butterfly needles require a waste tube to be drawn to clear line. Draw to fill line, short draws will be rejected.

Used for:

- Prothrombin (PT/INR)
- Prothrombin Time (PTT)
- D-Dimer
- Fibrinogen



Lavender – K2 EDTA

Last in order of draw, avoid K2 contamination to other samples.

Used for:

- CBC w/ Differential
- Hemogram and Platelet
- Hemoglobin A1C
- Sed Rate
- Reticulocyte



Helping all people live healthy lives

BD Vacutainer® Order of Draw for Multiple Tube Collections

Designed for Your Safety

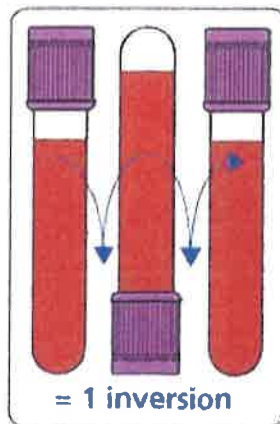
Reflects change in CLSI recommended Order of Draw (H3-A5, Vol 23, No 32, 8.10.2)

Closure Color	Collection Tube	Mix by Inverting
BD Vacutainer® Blood Collection Tubes (glass or plastic)		
	• Blood Cultures - SPS	8 to 10 times
	• Citrate Tube*	3 to 4 times
or	• BD Vacutainer® SST™ Gel Separator Tube	5 times
	• Serum Tube (glass or plastic)	5 times (plastic) none (glass)
	• BD Vacutainer® Rapid Serum Tube (RST)	5 to 6 times
or	• BD Vacutainer® PST™ Gel Separator Tube With Heparin	8 to 10 times
	• Heparin Tube	8 to 10 times
or	• EDTA Tube	8 to 10 times
	• BD Vacutainer® PPT™ Separator Tube K ₂ EDTA with Gel	8 to 10 times
	• Fluoride (glucose) Tube	8 to 10 times

* When using a winged blood collection set for venipuncture and a coagulation (citrate) tube is the first specimen tube to be drawn, a discard tube should be drawn first. The discard tube must be used to fill the blood collection set tubing's "dead space" with blood but the discard tube does not need to be completely filled. This important step will ensure proper blood-to-additive ratio. The discard tube should be a nonadditive or coagulation tube.

Note: Always follow your facility's protocol for order of draw

Handle all biologic samples and blood collection "sharps" (bancets, needles, luer adapters and blood collection sets) according to the policies and procedures of your facility. Obtain appropriate medical attention in the event of any exposure to biologic samples (for example, through a puncture injury) since they may transmit viral hepatitis, HIV (AIDS), or other infectious diseases. Utilize any built-in used needle protector if the blood collection device provides one. BD does not recommend resheating used needles, but the policies and procedures of your facility may differ and must always be followed. Discard any blood collection "sharps" in biohazard containers approved for their disposal.



BD Technical Services
1.800.631.0174
BD Customer Service
1.888.237.2762
www.bd.com/vacutainer

Urine Specimens

Patient Collection Instructions Included

STABILITY:

Urine samples must be refrigerated after collection, and delivered to the lab within 24 hours. Culture samples should be delivered as soon as possible. Samples should be labeled with name, DOB and collection time.



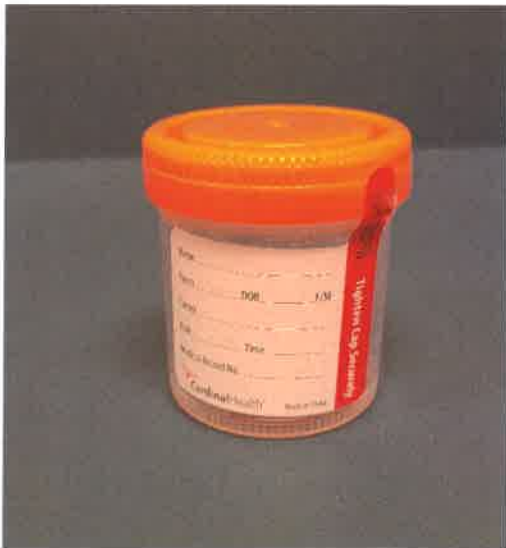
Clean Catch Collection

Materials:

Sterile collection cup, two (2) antiseptic towelettes

Used for:

- Urinalysis Routine
- Urinalysis with Reflex to Culture
- Urinalysis with Microscopic
- Urine Culture
- Urine Cytology



Dirty Catch Collection

Materials:

Sterile collection cup, collection hat (optional)

Used for:

- Chlamydia/GC Amplified Probe
(Must not have urinated in the last 60 minutes)
- MedTox/Drug Screening
- Nicotine/Metabolite Screen
- Creatinine/Albumin Ratio
- Urine Chemistries



24 Hour Collection

Materials:

2500 mL UrineGuard container, collection hat, patient collection instruction sheet (included)

Used for:

- 24-hour urine tests
- Metal-free jug required for certain 24-hour metal urine tests



CLEAN CATCH URINE COLLECTION

Technique for Women

1. Wash hands with soap and water.
2. Open and unfold the 2 towelette packages and open the screw cap container.
3. Separate the labia with one hand and hold them apart for the entire urine collection.
4. Cleanse the urinary opening with one of the towelettes. Make 3 passes using a front to back motion.
5. Repeat the process with the second towelette.
6. Void into the toilet for a couple seconds and then stop.
7. Restart the urine stream collection in the sterile container.
8. Without stopping the flow of urine, finish in the toilet.
9. Avoid touching the inside of the container. Screw the lid onto the container.

Techniques for Males

1. Wash hands thoroughly with soap and water.
2. Open and unfold the 2 towelette packages and open the screw cap container.
3. Cleanse the penis with one of the towelettes.
4. Repeat the process with the second towelette.
5. Void into the toilet for a couple seconds and then stop.
6. Restart the urine stream collection in the sterile container.
7. Without stopping the flow of urine, finish in the toilet.
8. Avoid touching the inside of the container. Screw the lid onto the container.

Copley Hospital Laboratory

Collection of 24 Hour Urine Specimens

PATIENT LABEL

REFRIGERATE SPECIMEN- no preservative is added

SPECIMEN COLLECTION:

ALWAYS REFRIGERATE SPECIMEN DURING COLLECTION

- 1) Patient should void first A.M. urine. DISCARD THIS SAMPLE. Record the date and time as "Collection Begun".
- 2) Collect all urine for the next 24 hours.
- 3) 24 hours after collection began, preferably the next morning, void and include this specimen. If unable to urinate at this time, obtain a sample as soon as possible. Record this time as "Collection Completed."
- 4) Collection must be 22-26 hours to be valid.

If you have any questions about the collection, call the Copley Lab at **888-8340**

Collection Begun : Date ___/___/___ Time: _____

Collection Finished: Date ___/___/___ Time: _____

Stool Specimens

Fecal Stability Chart Included



Unpreserved Specimens

Materials:

Sterile collection cup, collection hat, wooden tongue depressor or similar for transfer

Used for:

- H. pylori Antigen
- C Diff (Clostridium difficile)
- Fecal Occult Blood
- Calprotectin
- Lactoferrin
- Microsporidia
- Reducing Substances
- Fat Feces
- Pancreatic Elastase



Preserved Specimens – Black Top

Materials:

MCC Total-Fix Stool Collection Kit

- Add sample only to Fill Line

Used for:

- Ova and Parasite
- Giardia and Cryptosporidium antigens
- Giardia Microscopic exam
- Isospora/Cyclospora



Preserved Specimens – Orange Top

Materials:

Culture and Sensitivity Stool Transport Vial

- Add sample only to Fill Line

Used for:

- Fecal Bacterial Pathogens by PCR
- Fecal Culture Unusual Pathogens



Pinworm Collection

Materials:

Scientific Device Laboratory Pinworm Collector

Collection:

Follow instruction sheet provided with vial. Confirm expiration date prior to collection.

Fecal Stability Chart

Test	Stability	Sent to:
Fecal Bacterial pathogens by PCR, Feces culture Unusual pathogens	Raw specimen can be refrigerated in sterile container for up to 24 hours then must be put in orange top container. Orange top is stable for 4 days at Room temperature	UVMMMC
Ova and Parasite, Giardia and Cryptosporidium antigens, Isospora, Cyclospora	Raw specimen can be kept at Room Temp or refrigerated for up to 2 hours then must be put in a black container. Black container is stable for 3 days at Room Temp	UVMMMC
Giardia Microscopic exam (brushing)	Same as for Ova and Parasite (order O&P)	UVMMMC
H. pylori antigen	Stable for 3 days unpreserved and refrigerated. Must be frozen within 72 hours. Stable frozen for 60 days	UVMMMC
C Diff (Clostridium difficile)	Must be unpreserved specimen. Stable at room temp for 24 hours, refrigerated for 5 days	Copley
Occult blood	Raw specimens should be refrigerated until put on card. Inoculated card is stable for 14 days	Copley
Lactoferrin detection	Stable for 2 weeks refrigerated and unpreserved	Copley
Calprotectin	Stable for 3 days (72 hours) unpreserved and refrigerated. Must be frozen within 72 hours. Stable 7 days frozen.	Mayo
Microsporidia	Refrigerate unpreserved specimen. Stable for 7 days	Mayo
Reducing Substances	Must be unpreserved specimen. Must be frozen within 2 hours. Stable 7 days frozen.	Mayo
Fat Feces	Must be unpreserved specimen. Stable refrigerated 180 days. **Special patient preparation see Mayo FATF**	Mayo
Pancreatic Elastase	Stable for 3 days (72 hours) unpreserved and refrigerated. Must be frozen within 72 hours. Stable 28 days frozen.	Mayo

For "O&P Times 3" orders, only do Giardia Antigen ONCE

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STI Specimens

STABILITY:

Store collection kit at room temperature (15°C to 30°C).

Swabs stable in media at room temperature or refrigerated for 2 months, frozen at -20°C for 3 months



Endocervical/Urethral Specimens

Materials:

Hologic Aptima Unisex Swab Specimen Collection Kit for Endocervical and Male Urethral Swab Specimens

Used for:

- Female Endocervical Collection
- Male Urethral Collection



Vaginal and Other Source (Non-Traditional) Specimens

Materials:

Hologic Aptima Multitest Swab Specimen Collection Kit

Used for:

- Throat Collection
- Ocular Collection
- Rectal Collection
- Anal Collection

Culture Specimens

Store kits at room temperature.

Inoculated Culture Swabs are stable at room temperature or refrigerated up to 72 hours.

Avoid freezing or extremely high temperatures.



Materials:

Copan Transystem Sterile Transport Swab

Used for:

- Aerobic Cultures
- MRSA/MSSA Nasal Complete by PCR
- StrepScreen Culture (Group A Strep)
- Group B Strep DNA by PCR
- Pharyngitis Culture



Materials:

- Vacuette Viral Stabilization Tube

(vial may vary: M6 collection kit, M4RT, UTM, PBS and MedSchenker are acceptable)

- Anterior Nares swab

Used for:

- Non-Respiratory Viral PCR
 - o Herpes
 - o Varicella



Materials:

BBL CultureSwab Collection & Transport System

eSwab Transport System for Aerobic, Anaerobic & Fastidious Bacteria

Used for:

- UVM - Pertussis
- Mayo Clinic - Pertussis and Para-pertussis

COVID-19 Specimens

Differences in supply will result in a variety of COVID-19 transport media available. Always check expiration dates and storage requirements prior to use. Media vials must be labeled with the patient name and DOB.

COVID-19 Specimen Transport Media

Store at Room Temperature

Materials:

- Vacurette Virus Stabilization Tube
- Hardy Diagnostics Transport Medium
- Cepheid Xpert Viral Transport Medium
- Remel MicroTest M4RT Transport:
(two vial types)
 - o Large vial with red cap
 - o Small vial with purple cap

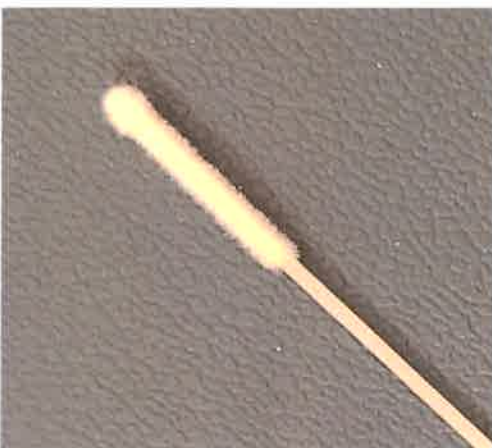
Used for:

- COVID-19
- Influenza
- RSV



Anterior Nares Swab (“Q-Tip”)

Have patient clear nostrils prior to swabbing. Swab 15 seconds in each nostril, to the depth of the nasal bone. Immediately place within transport medium, stable for 7 days refrigerated.



Nasopharyngeal Swab (“Pipe-cleaner”)

COVID-19 testing can be performed on either swab, but a Nasopharyngeal swab is **REQUIRED** when ordering Influenza/RSV. Influenza/RSV ordered on an Anterior Nasal swab will be rejected.

Have patient clear nostrils and tilt head back. Insert swab into one nostril until resistance and swab up to 30 seconds. Immediately place within transport medium, stable for 7 days refrigerated.